Peer

Responses to simulated nitrogen deposition by the neotropical epiphytic orchid *Laelia speciosa*

Edison A. Díaz-Álvarez^{1,2}, Roberto Lindig-Cisneros² and Erick de la Barrera²

¹ Posgrado en Ciencias Biológicas, Universidad Nacional Autónoma de México, Ciudad Universitaria, México, Distrito Federal, Mexico

² Instituto de Investigaciones en Ecosistemas y Sustentabilidad, Universidad Nacional Autónoma de México, Morelia, Michoacán, Mexico

ABSTRACT

Potential ecophysiological responses to nitrogen deposition, which is considered to be one of the leading causes for global biodiversity loss, were studied for the endangered endemic Mexican epiphytic orchid, Laelia speciosa, via a shadehouse dose-response experiment (doses were 2.5, 5, 10, 20, 40, and 80 kg N ha⁻¹ yr⁻¹) in order to assess the potential risk facing this orchid given impending scenarios of nitrogen deposition. Lower doses of nitrogen of up to 20 kg N ha yr^{-1} , the dose that led to optimal plant performance, acted as fertilizer. For instance, the production of leaves and pseudobulbs were respectively 35% and 36% greater for plants receiving 20 kg N ha yr⁻¹ than under any other dose. Also, the chlorophyll content and quantum yield peaked at 0.66 \pm 0.03 g m⁻² and 0.85 \pm 0.01, respectively, for plants growing under the optimum dose. In contrast, toxic effects were observed at the higher doses of 40 and 80 kg N ha yr⁻¹. The δ^{13} C for leaves averaged $-14.7 \pm 0.2\%$ regardless of the nitrogen dose. In turn, δ^{15} N decreased as the nitrogen dose increased from $0.9 \pm 0.1\%$ under 2.5 kg N ha⁻¹yr⁻¹ to $-3.1 \pm 0.2\%$ under 80 kg N ha⁻¹yr⁻¹, indicating that orchids preferentially assimilate NH_4^+ rather than NO_3^- of the solution under higher doses of nitrogen. Laelia speciosa showed a clear response to inputs of nitrogen, thus, increasing rates of atmospheric nitrogen deposition can pose an important threat for this species.

Subjects Ecology, Environmental Sciences, Plant Science, Soil Science, Coupled Natural and Human Systems

Keywords Acid rain, Biodiversity loss, CAM, Conservation physiology, δ^{15} N, Nitrogen pollution, Stable isotopes, Global change, Plant nutrition

INTRODUCTION

Anthropogenic atmospheric nitrogen deposition is considered among the leading global causes of biodiversity loss (*Vitousek, 1994*; *Chapin et al., 2000*; *Sala et al., 2000*). While nitrogen is an essential nutrient for all living organisms, its accelerated release to the atmosphere and ultimate deposition has caused saturation of various ecosystems around the world, leading to significant biodiversity loss by direct toxicity, acidification, and nutrient imbalances between nitrogen and other major nutrients (*Aber et al., 1989; Bauer*)

Submitted 17 February 2015 Accepted 24 May 2015 Published 23 June 2015

Corresponding author Erick de la Barrera, delabarrera@unam.mx

Academic editor Harpinder Sandhu

Additional Information and Declarations can be found on page 9

DOI 10.7717/peerj.1021

Copyright 2015 Díaz-Álvarez et al.

Distributed under Creative Commons CC-BY 4.0

OPEN ACCESS

et al., 2004; *Le Bauer & Treseder*, 2008; *Bobbink et al.*, 2010; *Templer, Pinder & Goodale*, 2012). Most studies regarding the effects of nitrogen deposition on biodiversity have been conducted in the USA and Europe, while studies from megadiverse countries are scant (*Bobbink et al.*, 2010). Considering that the latter countries tend to have developing economies and accelerated industrialization processes, it is urgent to determine the effects that current and future nitrogen deposition rates may have on their local biodiversities (*Austin et al.*, 2013).

A life-form particularly susceptible to the noxious effects of nitrogen deposition are epiphytic plants, such as certain species of orchids and bromeliads, given their reliance on atmospheric sources for nutrients and water (Zotz & Asshoff, 2010; Zotz et al., 2010; Mondragón, Valverde & Hernández-Apolinar, 2015). In this respect, Laelia speciosa (Kunth.) Shltr. (Orchidaceae) is an endemic, endangered orchid from central Mexico that has a cultural importance in Michoacán. Not only the plant is collected for its attractive flowers, but juice is extracted from its pseudobulbs and mixed with maize cane pith to produce a paste that is used for the production of sacred art in West Central Mexico (Soto-Arenas & Solano-Gómez, 2007). In addition to extractive pressure, this species faces environmental challenges considering that oak forests, to which this species is restricted, are likely to be severely reduced during the present century (Villers-Ruiz & Trejo-Vazquez, 2000; Rehfeldt et al., 2012). This study assessed whether nitrogen deposition can also pose a threat to this species. However, because current rates of nitrogen deposition are rather low within the area of distribution for L. speciosa (Díaz-Álvarez et al., 2014), it was deemed necessary to conduct a shadehouse dose-response experiment to determine the effects of potential future nitrogen deposition on this plant.

Indeed, the purpose of this study was to determine some ecophysiological responses of the endangered neotropical epiphytic orchid *Laelia speciosa* by means of a dose–response shadehouse experiment, in which, the organ production, chlorophyll content, chlorophyll fluorescence, carbon and nitrogen content and isotopic signatures were evaluated for assessing the potential risk that increasing rates of nitrogen deposition pose for this species.

MATERIALS AND METHODS

Plant material

Laelia speciosa is a sympodial epiphytic orchid with big and showy flowers that have pink to lilac-purple petals and a white lip. Flowers are produced during the spring, while an annually produced carbon-storing pseudobulb develops during the summer. *Laelia speciosa* grows in sub-humid temperate climates of central Mexico, between 1250 and 2500 m where its predominant phorophyte *Quercus deserticola* is also found (*Soto-Arenas & Solano-Gómez, 2007*).

Two-year old plants of *Laelia speciosa* obtained by *in vitro* propagation were transferred into 2L plastic pots containing tezontle (particles were 2–5 cm in diameter), a very porous volcanic rock that is extensively utilized for gardening and hydroponic horticulture given its suitable physicochemical properties (*Vargas-Tapia et al., 2008; Yañez-Ocampo et al., 2009*). Organic matter was removed from this tezontle by submersion in a SO₄H₂

aqueous solution (50% v/v) followed by a double rinse with deionized, distilled water. The procedure was repeated thrice. The pots were placed in a shadehouse for 2 years at Universidad Nacional Autónoma de México, Campus Morelia (19°38′55.9″N; 101°13′45″W; 1967 m, mean annual temperature of 18.3 °C, annual precipitation 773 mm; Servicio Meteorológico Nacional, 2011), where they were watered every other week until the start of the experiment. A total of 120 plants were selected at random and assigned to one of six groups, each with 20 plants, which received different doses of nitrogen. At the start of the experiment, the plants had 4 pseudobulbs and one leaf (15 cm in length).

Nitrogen deposition scenarios

Starting on 1 October 2011, the plants were watered weekly over two months with 50 ml of a modified Hoagland No. 2 solution from which the nitrogen was omitted to be able to simultaneously supply suitable amounts of nutrients and manipulate the dose of nitrogen dispensed to plants (Hoagland & Arnon, 1950; Nobel & de la Barrera, 2002). At the end of this period, six simulated atmospheric deposition scenarios were applied by adding 1, 4, 8, 16 or 32 mM of NH_4NO_3 to the watering solution, equivalent to 2.5, 10, 20, 40, or 80 kg of N ha⁻¹ yr⁻¹ respectively. In this respect, a threshold for plant physiological damage has been observed at 20 kg N ha⁻¹ yr⁻¹, while rates of 40 kg N ha⁻¹ yr⁻¹ are common in certain parts of Mexico City (Britto & Kronzucker, 2002; Secretaria del Medio Ambiente del Gobierno del Distrito Federal, 2012). The range of doses considered was chosen to establish the threshold for physiological damage for L. speciosa, and to determine the effects of deposition rates that are likely to occur during the present century. All amounts were calculated according to the area of the pot of 201 cm². Weekly applications of 50 ml of the experimental solutions were conducted over 26 weeks, from December 2011 to June 2012. This period corresponds to the growth season and reproductive development of Laelia speciosa (Halbinger & Soto, 1997; Soto-Arenas, 1994; Soto-Arenas & Solano-Gómez, 2007). Irrigation was carried out on the whole surface of the pot, the coarse substrate allowed the complete drainage of the nutrient solution, simulating what occurs in the canopy.

Physiological responses

Organ production

The emergence of flowers, which are displayed for a few weeks, was recorded weekly. In turn, the total production of new leaves and pseudobulbs, which are persistent, was recorded at the end of the experiment.

Chlorophyll content

Leaf discs were obtained with a cork borer (12-mm in diameter) from 5 plants per nitrogen deposition scenario to determine the concentration of chlorophyll a, chlorophyll b, and total chlorophyll in the plant tissue. The photosynthetic pigments were extracted by macerating leaf tissue with a chilled (3 °C) mortar and pestle in an aqueous solution of acetone (80% v/v) and brought to a final volume of 20 ml. Absorbance was measured at 663 and 646 nm with an EZ 301 spectrophotometer (Perkin Elmer, Waltham, Massachusetts, USA). Chlorophyll concentration was calculated following *Lichtenthaler (1987)*.

Chlorophyll fluorescence (Fv/Fm)

The maximum yield of the photosystem II (the ratio of variable to maximum fluorescence; Fv/Fm) was measured with an Opti-Science 05-30p Fluorometer (Hudson, New Hampshire, USA). Measurements were carried out before dawn, a common practice in plant ecophysiology (*Maxwell & Johnson, 2000*), for the leaves of five individuals per dose of nitrogen on 29 June and 2 July 2012.

Carbon and nitrogen content and isotopic composition

The leaves of plants growing under different nitrogen doses were collected on 4 July 2012 and dried at 80 °C in a gravity convection oven until reaching constant weight. This temperature has been found to be adequate for tropical succulents, whose membrane proteins can withstand temperatures that are substantially higher than for non-succulent species without incurring in damage (Nobel & de la Barrera, 2002; Drennan, 2009). The dried leaves were ground to a fine powder in a ball mill (Retsch MM300; Retsch, Vienna, Austria), wrapped into tin capsules (Costech Analytical, Inc., Valencia, California, USA), and weighed with a microbalance (0.01 mg, Sartorius, Göttingen, Germany). For each sample, the carbon and nitrogen content, as well as their isotopic proportions, were determined at the Stable Isotope Facility, University of Wyoming (Laramie, Wyoming, USA), with a Carlo Erba EA 1110 elemental analyzer (Costech Analytical Inc., Valencia, California, USA) attached to a continuous flow isotope ratio mass spectrometer (Finnigan Delta Plus XP, Thermo Electron Corp, Waltham, Massachusetts). Carbon and nitrogen isotope ratios, reported in parts per thousand, were calculated relative to the Vienna Pee Dee Belemnite (V-PDB) or atmospheric air standards, respectively. The analytical precision for δ^{13} C was $\pm 0.03\%$ (SD) and $\pm 0.06\%$ (SD) for δ^{15} N. The natural abundances of ¹³C and ¹⁵N were calculated as:

 $\delta^{13}C_{(\%_{0} \text{ versus V-PDB})} = (R_{\text{sample}}/R_{\text{standard}} - 1) \times 1000$ $\delta^{15}N_{(\%_{0} \text{ versus at-air})} = (R_{\text{sample}}/R_{\text{standard}} - 1) \times 1000$

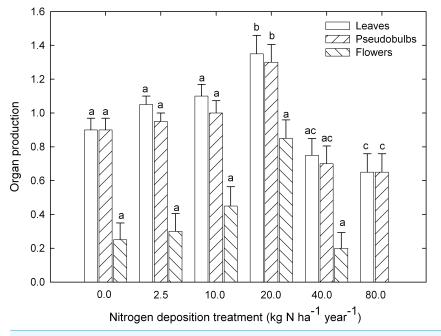
where, *R* is the ratio of ¹³C/¹²C for carbon and ¹⁵N/¹⁴N for nitrogen isotope abundance for a given sample (*Ehleringer & Osmond*, 1989; *Evans et al.*, 1996).

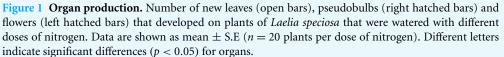
Statistical analyses

The effect of the simulated nitrogen deposition on organ production for *Laelia speciosa* was evaluated by means of a Kruskal-Wallis non-parametric ANOVA, because normality of data was not satisfied, followed by post-hoc Tukey tests ($P \le 0.05$). In turn, differences in the response of chlorophyll content, chlorophyll fluorescence, carbon and nitrogen content, and δ^{13} C and δ^{15} N, which achieved normality, were evaluated with a one-way ANOVA followed by the Holm-Sidak post-hoc test ($P \le 0.05$). All analyses were conducted on SigmaPlot 12 (Systat Software Inc., San Jose, California, USA).

Table 1 Statistical analyses. Kruskal-Wallis one-way ANOVA and parametric one-way ANOVA for the responses of *Laelia speciosa* individuals growing in a shadehouse under various rates of simulated nitrogen deposition.

	Response to nitrogen dose		
	d.f.	F	Р
Leaves	5	8.47	0.001
Pseudobulbs	5	7.04	0.001
Flowers	5	1.94	0.077
Total chlorophyll	5	15.68	0.001
Chla	5	6.67	0.001
Chlb	5	10.47	0.001
Fv/Fm	5	82.5	0.001
Carbon content	5	6.44	0.001
Nitrogen content	5	177.5	0.001
δ^{15} N	5	15.68	0.001
δ^{13} C	5	2.65	0.057





RESULTS

Organ production

After 26 weeks of watering the plants with different doses of nitrogen, the production of new organs was greater for those individuals that received 20 kg N ha⁻¹ yr⁻¹ than for those individuals receiving other nitrogen doses (Table 1; Fig. 1). In particular, 1.0 ± 0.1

leaves were produced per plant over the course of the experiment under most doses, except for the plants that received 20 kg N ha⁻¹ yr⁻¹, which produced 35% more leaves ($P \le 0.001$). Similar was the case for the 0.9 \pm 0.1 pseudobulbs produced per plant under most doses, except for the plants that received 20 kg N ha⁻¹ yr⁻¹, which produced 36% more pseudobulbs ($P \le 0.001$). In contrast, flowering was not significantly influenced by nitrogen dose (P = 0.077), with a production of 0.3 \pm 0.04 flowers per plant over the course of the experiment (Table 1; Fig. 1).

Chlorophyll fluorescence

The quantum efficiency of photosystem II (Fv/Fm) was similar among the groups of orchids that received up to 20 kg N ha⁻¹ yr⁻¹ amounting to 0.8, while a significant decrease of 23% was observed for plants irrigated with higher concentrations of nitrogen ($P \le 0.001$; Table 1; Fig. 2A).

Chlorophyll content

Total chlorophyll content for the leaf tissue of *Laelia speciosa* increased as the nitrogen dose increased, peaking at 0.7 ± 0.0 g m⁻² for plants irrigated with 20 kg N ha⁻¹ yr⁻¹ ($P \le 0.001$; Table 1; Fig. 2B), while the higher doses of nitrogen resulted in a 38% reduction of the pigment. Similarly, the chlorophyll a concentration of 0.5 ± 0.4 g m⁻² was the maximum for plants growing under 20 kg N ha⁻¹yr⁻¹, and it was 30% lower under all other nitrogen doses ($P \le 0.001$). In turn, chlorophyll b did not respond to nitrogen, averaging 0.1 ± 0.0 g m⁻² regardless of the dose under which plants grew (Table 1; Fig. 2B).

Carbon and nitrogen content and isotopic composition

The carbon content of *Laelia speciosa* increased with the nitrogen dose peaking at $46.1 \pm 0.3\%$ (dry mass basis) at 20 and 40 kg N ha⁻¹ yr⁻¹ and decreased to $45.2 \pm 0.3\%$ at 80 kg N ha⁻¹ yr⁻¹ ($P \le 0.001$; Table 1; Fig. 2C).

The nitrogen content for *Laelia speciosa* also increased with the nitrogen dose. For the plants that received up to 10 kg N ha⁻¹ yr⁻¹ the nitrogen content averaged $1.2 \pm 0.0\%$ (dry mass basis), reaching $2.4 \pm 0.0\%$ at 80 kg N ha⁻¹ yr⁻¹ ($P \le 0.001$; Table 1; Fig. 2D).

The δ^{13} C for leaves of *Laelia speciosa* averaged $-14.7 \pm 0.2\%$ and did not change with the nitrogen dose (P = 0.057; Table 1). In contrast, the leaf δ^{15} N significantly decreased at higher nitrogen doses. The δ^{15} N averaged $0.9 \pm 0.1\%$ for plants that received up to 10 kg N ha⁻¹ yr⁻¹, a δ^{15} N similar to the δ^{15} N of $1.1 \pm 0.1\%$ measured for the NH₄NO₃ utilized for the nutrient solution. The higher doses of nitrogen led to significant decreases of δ^{15} N, reaching the minimum of $-3.1 \pm 0.2\%$ for plants growing under 80 kg N ha⁻¹ yr⁻¹ ($P \le 0.001$; Table 1; Fig. 2E).

DISCUSSION

An intermediate nitrogen dose of 20 kg N ha⁻¹ yr⁻¹ was the most favorable for the production of new organs by *Laelia speciosa*. Lower doses did not improve plant development substantially but higher doses were inhibiting. In this respect, while nitrogen availability may increase leaf production and growth, large quantities of nitrogen limit the

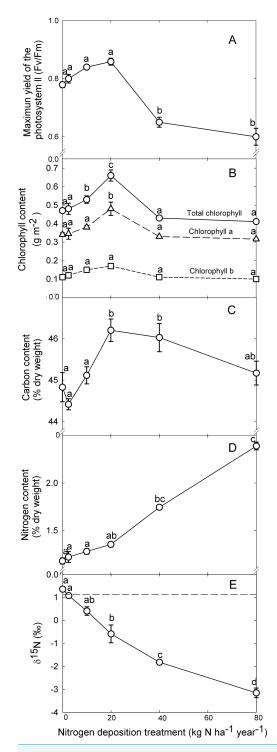


Figure 2 Ecophysiological responses for leaves of *L. speciosa* to simulated nitrogen deposition. (A) Ratio of variable to maximum chlorophyll fluorescence; (B) Tissue content (area basis) for total chlorophyll (circles), chlorophyll-a (triangles), and chlorophyll-b (square); (C) Carbon and (D) nitrogen content (dry mass basis) and (E) δ^{15} N. Data are shown as mean \pm S.E. (n = 5 plants per dose of nitrogen). For each panel, different letters indicate significant differences (P < 0.05).

availability of other nutrients, restricting the plant's ability to produce foliar mass (*Evans*, 1989; Asner, Seastedt & Townsend, 1997; Aber et al., 1998; Sánchez et al., 2000; Zotz & Asshoff, 2010; Díaz-Álvarez et al., 2014). Such behavior was observed for Laelia speciosa that showed a substantial reduction in the production of new organs, suggesting noxious effects of the simulated nitrogen deposition. The effect of nitrogen fertilization on *Cymbidium* hybrids is an increased pseudobulb production (*Barman et al., 2004*). In turn, pseudobulb growth for *Dendrobium nobile* peaks at nitrogen doses of 1.9 mM (*Bichsel, Starman & Yin-Tung, 2008*).

Total chlorophyll content is proportional to the content of nitrogen in leaves, which typically ranges between 0.4 and 0.5 g m⁻² (*Evans, 1989; Nobel, 1999; Nobel & de la* Barrera, 2002). Indeed, for Laelia speciosa, chlorophyll content increased with the dose of nitrogen, suggesting that this plant was able to assimilate and utilize the supplied nitrogen for the production of photosynthetic pigments. However, the higher doses also resulted in a drastic decrease of the chlorophyll content, as has been documented for other plant species (Baxter, Emes & Lee, 1992; Majerowicz et al., 2000; Lin et al., 2007; Arróniz-Crespo et al., 2008; Ying-Chun et al., 2010). Such a decrease in the chlorophyll content can be explained by the resulting imbalance of the nitrogen to magnesium ratio in the leaf (Nakaji et al., 2001; Wortman et al., 2012). Excessive nitrogen in the cell promotes release of protons (H^+) and accumulation of phenolic compounds and hydrogen peroxide, as a result, the pH can be altered impeding chlorophyll production and loss of Mg²⁺ (Mangosá & Berger, 1997; Sánchez et al., 2000; Britto & Kronzucker, 2002). Changes in chlorophyll content for Laelia speciosa were accompanied by changes in the efficiency of photosystem II, which can be attributed to oxidative stress in the thylakoids that results in the blockage of electron transport to the oxidation site, as a consequence of low available energy for photosynthesis (Maxwell & Johnson, 2000; Poorter, 2000; Hogewoning & Harbinson, 2007; Lichtenthaler et al., 2007; Baker, 2008; Calatayud et al., 2008; Guidi & Degl'Innocenti, 2008; Massacci et al., 2008).

Plants tend to increase their rates of carbon fixation when nitrogen is added (*Brown* et al., 1996; Bauer et al., 2004; Le Bauer & Treseder, 2008). However, under conditions of chronic nitrogen additions the photosynthetic capacity is inhibited because most of the excess nitrogen is not invested into the primary processes of carboxylation (*Brown et al., 1996; Bauer et al., 2004*). This also causes an increase and later reduction in carbon content for plants subjected to increasing doses of nitrogen, as was observed here for *Laelia speciosa*. However, the observed δ^{13} C values for *Laelia speciosa* which were within the range for CAM plants, did not change under the different nitrogen doses utilized, contrasting with δ^{13} C measured for C₃ plants subjected to supplementary nitrogen that became increasingly negative (*Raven & Farquhar, 1990; Magalhaes, Huber & Tsai, 1992*).

Isotopic discrimination against ¹⁵N increases in plants as the nitrogen availability increases because its assimilation is more energetically costly than for the more abundant ¹⁴N. This so called isotopic effect results in δ^{15} N values of the product that are lower than those of the substrate (*Evans, 2001; Kolb & Evans, 2003; Ariz et al., 2011*). The observed discrimination against ¹⁵N for *Laelia speciosa* leaves has also been observed

for various species, such as *Oryza sativa, Pinus sylvestris*, and *Trapa japonica*, species which discriminate between 0.9 and 13^{\%} when supplied with increasing doses of nitrogen in form of NH⁺₄ (*Yoneyama et al., 1991*; *Högberg et al., 1999*; *Yoneyama et al., 2001*; *Maniruzzaman & Asaeda, 2012*). When the nitrogen source is NH⁺₄, this compound is directly assimilated by the plant cell as amino acids and the involved enzyme, glutamine-synthetase, can discriminate up to 17^{\%}. On the contrary, plants watered with NO⁻₃ have positive δ^{15} N values that have been associated with nitrogen lost via root efflux and exudates or loss of NH³ through the stomata, processes that favor the lighter isotope (*O'Deen, 1989*; *Yoneyama et al., 2001*; *Ariz et al., 2011*).

Laelia speciosa showed a clear response to increasing doses of nitrogen. Doses of up to 20 kg N ha⁻¹ year⁻¹ enhanced its physiological performance, while higher doses were toxic. The rates of nitrogen deposition in México, where *Laelia speciosa* is endemic, could exceed 25 kg N ha⁻¹ year⁻¹ by mid-century (*Galloway et al., 2004*; *Phoenix et al., 2006*; *Galloway et al., 2008*). As a result, nitrogen deposition poses an actual threat for the persistence of this endangered species as other components of global change represent for many other epiphytic vascular plants (*Zotz et al., 2010*; *Mondragón, Valverde & Hernández-Apolinar, 2015*). Future works should consider the effects of nitrogen deposition on wild populations of this and other tropical epiphytic plants. A better understanding of the effects of increasing nitrogen deposition from human activities is of urgent importance, as species ecophysiological response, as those studied here, may be affected, with potentially negative consequences in ecosystem biodiversity and function.

ACKNOWLEDGEMENTS

We thank Dr. I Ávila Díaz for providing the plants utilized in this study; the personnel of the IIES Botanical Gardens for maintenance of the plants before the experiment, especially Ms. MD Lugo and Mr. J. Martinez Cruz; and Dr. DG Williams at the University of Wyoming for his guidance during the elemental and isotopic analyses of the samples.

ADDITIONAL INFORMATION AND DECLARATIONS

Funding

We thank funding by Dirección General del Personal Académico (PAPIIT IN224910 and RN204013), UNAM and institutional funds from Instituto de Investigaciones en Ecosistemas y Sustentabilidad (IIES), UNAM. EADA was funded by a generous graduate research fellowship from Consejo Nacional de Ciencia y Tecnología, México. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Grant Disclosures

The following grant information was disclosed by the authors: Dirección General del Personal Académico: PAPIIT IN224910, RN204013. Instituto de Investigaciones en Ecosistemas y Sustentabilidad (IIES), UNAM. Consejo Nacional de Ciencia y Tecnología, México.

Competing Interests

The authors declare there are no competing interests.

Author Contributions

- Edison A. Díaz-Álvarez conceived and designed the experiments, performed the experiments, analyzed the data, wrote the paper, prepared figures and/or tables, reviewed drafts of the paper.
- Roberto Lindig-Cisneros conceived and designed the experiments, contributed reagents/materials/analysis tools, reviewed drafts of the paper, supervised plant nutrition experiments.
- Erick de la Barrera conceived and designed the experiments, contributed reagents/materials/analysis tools, wrote the paper, prepared figures and/or tables, reviewed drafts of the paper.

Supplemental Information

Supplemental information for this article can be found online at http://dx.doi.org/ 10.7717/peerj.1021#supplemental-information.

REFERENCES

- Aber JD, McDowell W, Nadelhoffer KJ, Magill A, Berntso GM, Kamakea M, McNulty S, Currie W, Rustad L, Fernández I. 1998. Nitrogen saturation in temperate forest ecosystems—hypothesis revisited. *Bioscience* 48:921–934 DOI 10.2307/1313296.
- Aber JD, Nadelhoffer KJ, Steudler P, Melillo JM. 1989. Nitrogen saturation in northern forest ecosystems. *Bioscience* 39:378–386 DOI 10.2307/1311067.
- Ariz I, Cruz C, Moran JF, González-Moro MB, García-Olaverri C, González-Murua C, Martins-Loução MA, Aparicio-Trejo PM. 2011. Depletion of the heaviest stable N isotope is associated with NH₄⁺/NH₃ toxicity in NH₄⁺-fed plants. *BMC Plant Biology* 2011(11):83 DOI 10.1186/1471-2229-11-83.
- Arróniz-Crespo M, Leake JR, Horton P, Phoenix GK. 2008. Bryophyte physiological responses to, and recovery from, long-term nitrogen deposition and phosphorus fertilization in acidic grassland. *New Phytologist* 180:864–874 DOI 10.1111/j.1469-8137.2008.02617.x.
- Asner GP, Seastedt TR, Townsend AR. 1997. The decoupling of terrestrial carbon and nitrogen cycles. *Bioscience* 47:226–234 DOI 10.2307/1313076.
- Austin AT, Bustamante MMC, Nardoto GB, Mitre SK, Pérez T, Ometto JPHB, Ascarrunz NL, Forti MC, Longo K, Gavito ME, Enrich-Prast A, Martinelli LA. 2013. Latin America's nitrogen challenge. *Science* 340:149 DOI 10.1126/science.1231679.
- **Baker NR. 2008.** Chlorophyll fluorescence: a probe of photosynthesis in vivo. *Annual Review of Plant Biology* **59**:89–113 DOI 10.1146/annurev.arplant.59.032607.092759.
- Barman P, Rajni K, Naik SK, Upadhyaya RC. 2004. Effect of nitrogen, phosphorus and potassium on growth and flowering of *Cimbidiu. Journal of Ornamental Horticulture* 7:235–238.
- Bauer GA, Bazzaz FA, Minocha R, Long S, Magill A, Aber J, Berntson GM. 2004. Effects of chronic N additions on tissue chemistry, photosynthetic capacity, and carbon sequestration potential of a red pine (*Pinus resinosa* Ait.) stand in the NE United States. *Forest Ecology and Management* 196:173–186 DOI 10.1016/j.foreco.2004.03.032.

- Baxter R, Emes MJ, Lee JA. 1992. Effects of an experimentally applied increase in ammonium on growth and amino-acid metabolism of Sphagnum cuspidatum Ehrh. ex. Hoffm. from differently polluted areas. *New Phytologist* 120:265–274 DOI 10.1111/j.1469-8137.1992.tb05663.x.
- **Bichsel RB, Starman W, Yin-Tung W. 2008.** Nitrogen, phosphorus, and potassium requirements for optimizing growth and flowering of the nobile dendrobium as a potted orchid. *HortScience* **43**:328–332.
- Bobbink R, Hicks K, Galloway J, Spranger T, Alkemade R, Ashmore M, Bustamante M, Cinderby S, Davidson E, Dentener F, Emmett B, Erisman JW, Fenn M, Gilliam F, Nordin A, Pardo L, De Vries W. 2010. Global assessment of nitrogen deposition effects on terrestrial plant diversity: a synthesis. *Ecological Applications* 20:30–59 DOI 10.1890/08-1140.1.
- **Britto T, Kronzucker H. 2002.** NH₄⁺ toxicity in higher plants: a critical review. *Journal of Plant Physiology* **159**:567–584 DOI 10.1078/0176-1617-0774.
- Brown KR, Thompson WA, Camm EL, Hawkins BJ, Guy RD. 1996. Effects of N addition rates on the productivity of *Picea Sitchensis, Thuja plicata*, and *Tsuga heterophylla* seedlings: II. Photosynthesis, ¹³C discrimination and N partitioning in foliage. *Trees* 10:198–205 DOI 10.1007/BF02340772.
- **Calatayud A, Gorbe E, Roca D, Martínez PF. 2008.** Effect of two nutrient solution temperatures on nitrate uptake, nitrate reductase activity, NH₄⁺ concentration and chlorophyll a fluorescence in rose plants. *Environmental and Experimental Botany* **64**:65–74 DOI 10.1016/j.envexpbot.2008.02.003.
- Chapin III FS, Zavaleta ES, Eviner VT, Naylor RL, Vitousek PM, Reynolds HL, Hooper DU, Lavorel S, Sala OE, Hobbie SE, Mack MC, Díaz S. 2000. Consequences of changing biodiversity. *Nature* 405:234–242 DOI 10.1038/35012241.
- Díaz-Álvarez EA, Lindig-Cisneros R, Reyes-García C, de la Barrera E. 2014. Effects of simulated nitrogen deposition and a stable isotopic assessment for the neotropical epiphytic orchid *Laelia speciosa*. *PeerJ PrePrint* 2:e771v1.
- **Drennan PM. 2009.** Temperature influences on plant species of arid and semi-arid regions with emphasis on CAM succulents. In: de la Barrera E, Smith WK, eds. *Perspectives in biophysical plant ecophysiology: a tribute to Park S. Nobel.* México City: Universidad Nacional Autonoma de México, 57–94.
- **Ehleringer JR, Osmond BO. 1989.** Stable isotopes. In: Pearcy RW, Ehleringer JR, Mooney HA, Rundel PW, eds. *Plant physiological ecology*. London: Chapman & Hall.
- Evans RD, Bloom AJ, Sukrapanna SS, Ehleringer JR. 1996. Nitrogen isotope composition of tomato (*Lycopersicon esculentum* Mill. cv. T-5) grown under ammonium or nitrate nutrition. *Plant Cell and Environment* 19:1317–1323 DOI 10.1111/j.1365-3040.1996.tb00010.x.
- **Evans RD. 2001.** Physiological mechanisms influencing plant nitrogen isotope composition. *Trends in Plant Science* **6**:121–126 DOI 10.1016/S1360-1385(01)01889-1.
- **Evans JR. 1989.** Photosynthesis and nitrogen relationships in leaves of C₃ plants. *Oecologia* **78**:9–19 DOI 10.1007/BF00377192.
- Galloway JN, Townsend AR, Erisman JW, Bekunda M, Cai Z, Freney JR, Martinelli LA, Seitzinger SP, Sutton MA. 2008. Transformation of the nitrogen cycle: recent trends, questions, and potential solutions. *Science* 320:889–892 DOI 10.1126/science.1136674.
- Galloway JN, Dentener FJ, Capone DG, Boyer EW, Howarth RW, Seitzinger SP, Asner GP, Cleveland CC, Green PA, Holland EA, Karl DM, Michaels AF, Porter JH, Townsend AR, Vörösmarty CJ. 2004. Nitrogen cycles: past, present, and future. *Biogeochemistry* 70:153–226 DOI 10.1007/s10533-004-0370-0.

- Guidi L, Degl'Innocenti E. 2008. Ozone effects on high light-induced photoinhibition in *Phaseolus vulgaris*. *Plant Science* 174:590–596 DOI 10.1016/j.plantsci.2008.03.003.
- Halbinger F, Soto MA. 1997. Laelias of Mexico. Revista del Herbario AMO 15:1-160.
- Hoagland DR, Arnon DI. 1950. The culture method for growing plants without soil. *California Agriculture Experimental Circle* 347:1–32.
- Högberg P, Högberg MN, Quist ME, Ekblad A, Näsholm T. 1999. Nitrogen isotope fractionation during nitrogen uptake by ectomycorrhizal and nonmycorrhizal *Pinus sylvestris*. *New Phytologist* 142:569–576 DOI 10.1046/j.1469-8137.1999.00404.x.
- **Hogewoning SW, Harbinson J. 2007.** Insights on the development, kinetics, and variation of photoinhibition using chlorophyll fluorescence imaging of a chilled, variegated leaf. *Journal of Experimental Botany* **58**:453–463 DOI 10.1093/jxb/erl219.
- Kolb K, Evans R. 2003. Influence of nitrogen source and concentration on nitrogen isotopic discrimination in two barley genotypes (*Hordeum vulgare L.*). *Plant Cell and Environment* 26:1431–1440 DOI 10.1046/j.1365-3040.2003.01066.x.
- Le Bauer DS, Treseder KK. 2008. Nitrogen limitation of net primary productivity in terrestrial ecosystems is globally distributed. *Ecology* **89**:371–379 DOI 10.1890/06-2057.1.
- Lichtenthaler HK. 1987. Chlorophylls and carotenoids: pigments of photosynthetic membranes. Methods in Enzymology 148:350–382.
- Lichtenthaler HK, Ač A, Marek MV, Kalina J, Urban O. 2007. Differences in pigment composition, photosynthetic rates and chlorophyll fluorescence images of sun and shade leaves of four tree species. *Plant Physiology Biochemistry* **45**:577–588 DOI 10.1016/j.plaphy.2007.04.006.
- Lin Y, Duan L, Yang YS, Zhao DW, Zhang DB, Hao JM. 2007. Contribution of simulated nitrogen deposition to forest soil acidification in area with high sulfur deposition. *Environmental Science* 28:640–646.
- Magalhaes JR, Huber DM, Tsai CY. 1992. Evidence of increased ¹⁵N ammonium assimilation in tomato plants with exogenous a-ketoglutarate. *Plant Science* **85**:135–141 DOI 10.1016/0168-9452(92)90108-X.
- Majerowicz N, Kerbauy GB, Nievola CC, Suzuki RM. 2000. Growth and nitrogen metabolism of *Catasetum fimbriatum* (Orchidaceae) grown with different nitrogen sources. *Environmental and Experimental Botany* 44:195–206 DOI 10.1016/S0098-8472(00)00066-6.
- Mangosá TJ, Berger RG. 1997. Determination of major chlorophyll degradation products. *European Food Research and Technology* 204:345–350.
- Maniruzzaman M, Asaeda T. 2012. Fractionation of stable isotopes of nitrogen in *Trapa japonica* during uptake of NH₄⁺-N. *Research in Plant Biology* **2**:48–52.
- Massacci A, Nabiev SM, Pietrosanti L, Nematov SK, Chernikova TN, Thor K, Leipner J. 2008. Response of the photosynthetic apparatus of cotton (Gossypium hirsutum) to the onset of drought stress under oak forest conditions studied by gas exchange analysis and chlorophyll fluorescence imaging. *Plant Physiology Biochemistry* **46**:189–195 DOI 10.1016/j.plaphy.2007.10.006.
- Maxwell K, Johnson G. 2000. Chlorophyll fluorescence—a practical guide. *Journal of Experimental Botany* 51:659–668 DOI 10.1093/jexbot/51.345.659.
- Mondragón D, Valverde T, Hernández-Apolinar M. 2015. Population ecology of epiphytic angiosperms: a review. *Tropical Ecology* 56:01–39.

- Nakaji T, Fukami M, Dokiya Y, Izuta T. 2001. Effects of high nitrogen load on growth, photosynthesis and nutrient status of *Cryptomeria japonica* and *Pinus densiflora* seedlings. *Trees* 15:453–461 DOI 10.1007/s00468-001-0130-x.
- **Nobel PS. 1999.** *Physicochemical and environmental plant physiology.* 2nd edition. San Diego California: Academic Press.
- Nobel PS, de la Barrera E. 2002. Nitrogen relations for net CO₂ uptake by the cultivated hemiepiphytic cactus, *Hylocerus undatus*. *Scientia Horticulturae* 96:281–292 DOI 10.1016/S0304-4238(02)00060-2.
- **O'Deen WA. 1989.** Wheat volatilized ammonia and resulting nitrogen isotopic fractionation. *Agronomy Journal* **81**:980–985 DOI 10.2134/agronj1989.00021962008100060027x.
- Phoenix GK, Hicks WK, Cinderby S, Kuylenstierna CI, Stock WD, Dentener FJ, Giller KE, Austin AT, Lefroy DB, Gimeno BS, Ashmore MR, Ineson P. 2006. Atmospheric nitrogen deposition in world biodiversity hotspots: the need for a greater global perspective in Assessing N deposition impacts. *Global Change Biology* 12:470–476 DOI 10.1111/j.1365-2486.2006.01104.x.
- **Poorter HN. 2000.** The role of biomass allocation in the growth response of plants to different level of light, CO₂, nutrients and water: a quantitative review. *Australian Journal of Plant Physiology* **27**:597–607 DOI 10.1071/PP99173_CO.
- Raven JA, Farquhar GD. 1990. The influence of N metabolism and organic acid synthesis on the natural abundance of isotopes of carbon in plants. *New Phytologist* 116:505–529 DOI 10.1111/j.1469-8137.1990.tb00536.x.
- Rehfeldt GE, Crookston NL, Sáenz-Romero C, Campbell EM. 2012. North American vegetation model for land-use planning in a changing climate: a solution to large classification problems. *Ecological Applications* 22:119–141 DOI 10.1890/11-0495.1.
- Sala OE, Chapin III SF, Armesto JJ, Berlow E, Bloomfield J, Dirzo R, Huber-Sanwald E, Huenneke LF, Jackson RB, Kinzig A, Leemans R, Lodge DM, Mooney HA, Oesterheld M, Poff NL, Sykes MT, Walker BH, Walker M, Wal DH. 2000. Global biodiversity scenarios for the year 2100. *Science* 287:1770–1774 DOI 10.1126/science.287.5459.1770.
- Sánchez E, Soto J, García PC, López-Lefebre L, Rivero R, Ruíz JM, Romero L. 2000. Phenolic compounds and oxidative metabolism in green vean plants under nitrogen toxicity. *Australian Journal of Plant Physiology* 27:973–978.
- Secretaria del Medio Ambiente del Gobierno del Distrito Federal. 2012. Calidad del aire en la Ciudad de México Informe 2011. México City: SEMARNAT. 164 pp. Available at http://www.sedema.df.gob.mx/flippingbook/informe_anual_calidad_aire_2011/.
- Soto-Arenas M. 1994. Population studies in Mexican orchids. In: Pridgeon AM, ed. Proceedings of the 14th world orchid conference. Glasgow: HMSO Publ.
- **Soto-Arenas MA, Solano-Gómez AR. 2007.** Ficha técnica de *Laelia speciosa*. In: Soto-Arenas MA, ed. *Información actualizada sobre las especies de orquídeas del PROY-NOM-059-ECOL-2000.* México. D.F: Instituto Chinoin A.C., Herbario de la Asociación Mexicana de Orquideología A.C. Bases de datos SNIB-CONABIO. Proyecto No. W029.
- Templer PH, Pinder RW, Goodale CL. 2012. Effects of nitrogen deposition on greenhouse-gas fluxes for forests and grasslands of North America. *Frontiers in Ecology and Environment* 10:547–553 DOI 10.1890/120055.
- Vargas-Tapia P, Castellanos–Ramos JZ, Muñoz–Ramos JJ, Sánchez–García P, Tijerina–Chávez L, López–Romero RM, Martínez–Sánchez C, Ojodeagua–Arredondo JL.
 2008. Effect of particle size on some physical properties of tezontle (volcanic rock) from the State of Guanajuato. *Agricultura técnica en México* 34:323–331.

- Villers-Ruiz L, Trejo-Vazquez I. 2000. El cambio climático y la vegetación en México. In: Gay-García C, ed. México: una visión hacia el siglo XXI. El cambio climático en México. Instituto Nacional de Ecología. Universidad Nacional Autónoma de México, US Country Studies Program, 57–66.
- Vitousek PM. 1994. Beyond global warming: ecology and global chance. *Ecology* 75:1861–1876 DOI 10.2307/1941591.
- Wortman E, Tomaszewski T, Waldner P, Schleppi P, Thimonier A, Eugster W, Buchmann N, Sievering H. 2012. Atmospheric nitrogen deposition and canopy retention influences on photosynthetic performance at two high nitrogen deposition Swiss forests. *Tellus B*. 64:17216 DOI 10.3402/tellusb.v64i0.17216.
- Yañez-Ocampo G, Sanchez-Salinas E, Jimenez-Tobonb GA, Michel Penninckx M, Ortíz-Hernández ML. 2009. Removal of two organophosphate pesticides by a bacterial consortium immobilized in alginate or tezontle. *Journal of Hazardous Materials* 168:1554–1561 DOI 10.1016/j.jhazmat.2009.03.047.
- Ying-Chun L, Hou-Bao F, Yan-Yan L, Wen-Fei L, Ying-Hong Y. 2010. Effects of simulated nitrogen deposition on growth and photosynthesis of 1-year-old Chinese fir (*Cunninghamia lanceolata*) seedlings. *Acta Ecologica Sinica* **30**:150–154 DOI 10.1016/j.chnaes.2010.04.005.
- Yoneyama T, Omata T, Nakata S, Yazaki J. 1991. Fractionation of nitrogen isotopes during the uptake and assimilation of ammonia by plants. *Plant Cell Physiology* **32**:1211–1217.
- Yoneyama T, Matsumaru T, Usui K, Engelaar WMHG. 2001. Discrimination of nitrogen isotopes, during absorption of ammonium and nitrate at different nitrogen concentrations by rice (*Oryza sativa* L.) plants. *Plant Cell and Environment* 24:133–139 DOI 10.1046/j.1365-3040.2001.00663.x.
- Zotz G, Asshoff R. 2010. Growth in epiphytic bromeliads: response to the relative supply of phosphorus and nitrogen. *Plant Biology* 12:108–113 DOI 10.1111/j.1438-8677.2009.00216.x.
- Zotz G, Bogusch W, Hiezt P, Ketteler N. 2010. Growht of epiphytic bromeliads in a changing world: the effects of CO₂, water and nutrient supply. *Acta Oecologica* 36:659–665 DOI 10.1016/j.actao.2010.10.003.